GALENVS

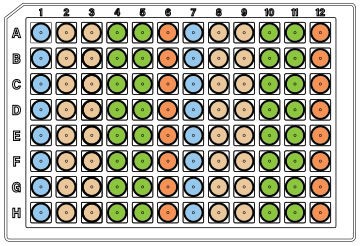
magnetic

Plant RNA Extraction Kit

Quick Start Guide

Part A RNA Extraction

- Binding Buffer (800µl) — Columns 1 & 7
- Wash #2 Buffer (600µl) — Columns 4 & 10, 5 & 11
- Wash #1 Buffer (600µl) — Columns 2 & 8, 3 & 9
- Elution Buffer (100µl) — Columns 6 & 12



200µl sample + 40µl Binding Magnetic Nanoparticles added to Binding Buffer

100µl extracted and purified RNA elution

200µl sample + 40µl Binding Magnetic Nanoparticles added to Binding Buffer

100µl extracted and purified RNA elution

miQron Protocol Update

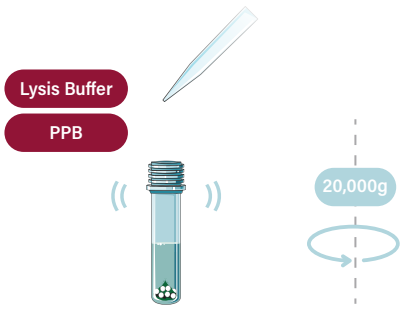
To import the updated protocol into the miQron, press the **Scan Protocol** icon from the **Run** menu (protocol list view window). Use the scanner on the QR code below.

PRKIT-A PROTOCOL V1.0

1 Add up to 50mg of fresh plant leaves sample to the lysis bead tube provided.

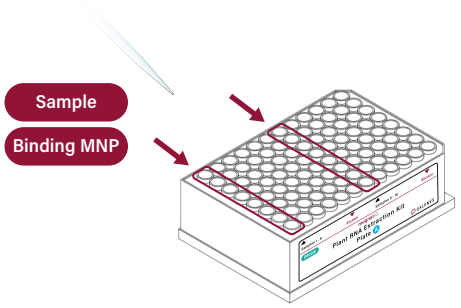
2 Add 600µl Lysis Buffer, and 60µl PPB.

Mix for 10 mins using TissueLyser at max speed or vortex for 10 mins; then centrifuge at 20,000g for 2 mins.

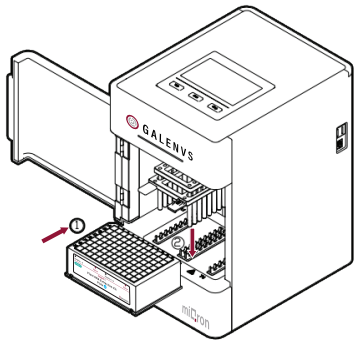


3 To the **RNA Extraction Kit Plate** A transfer 200µl of supernatant to Binding Buffer #1 (columns 1 & 7). Then add 40µl of Binding Magnetic Nanoparticles.

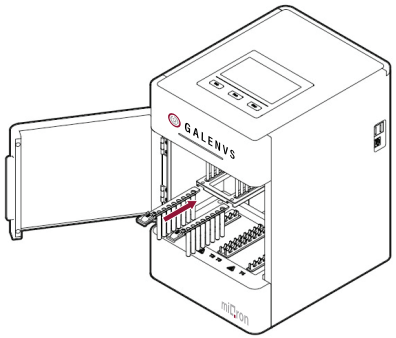
You can add up to 16 samples.



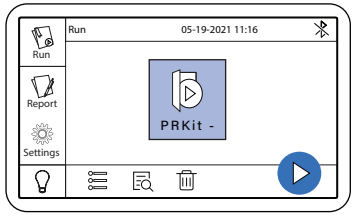
4 Place plate into the miQron, taking care that the label is facing outward.



5 Insert two combs.



6 Select **PRKit - Part A** and press ▶



When program is complete, remove plate from miQron and discard combs.

Proceed to Part B DNase Treatment

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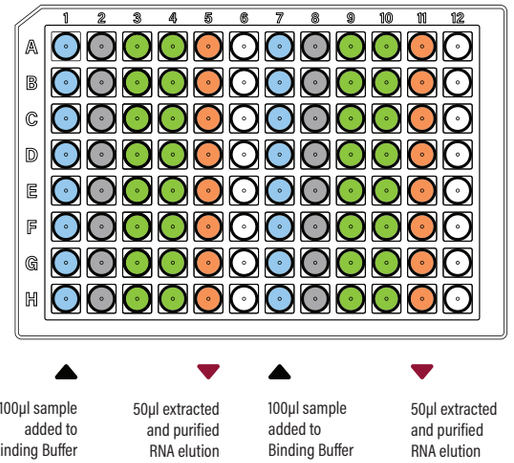
Plant RNA Extraction Kit

Quick Start Guide

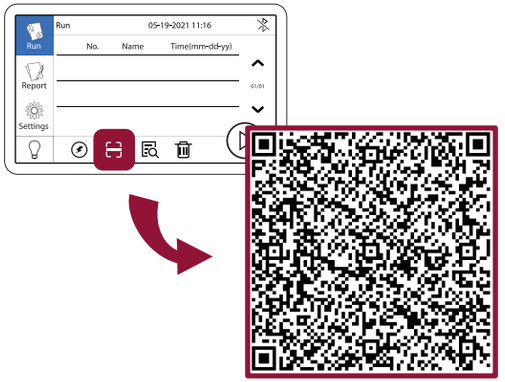
Part B DNase Treatment



- Binding Buffer (400µl) — Columns 1 & 7
- Functionalized Beads (200µl) — Columns 2 & 8
- Wash #3 Buffer (600µl) — Columns 3 & 9, 4 & 10
- Elution Buffer (50µl) — Columns 5 & 11

miQron Protocol Update

To import the updated protocol into the miQron, press the **Scan Protocol** icon from the **Run** menu (protocol list view window). Use the scanner on the QR code below.

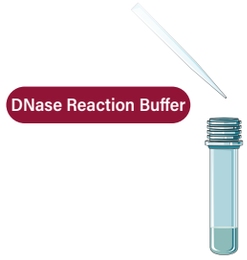


PRKIT-B PROTOCOL V1.0

7 Add 500µl of DNase Reaction Buffer to DNase pellet.

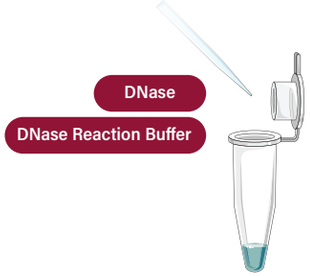
Mix by gently inverting the tube.

Reconstituted pellet must be stored at -20 °C.

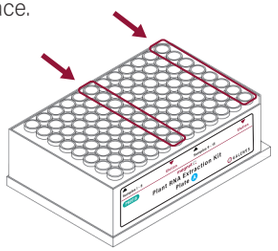


8 For each RNA sample, add 10µl of the reconstituted DNase prepared in the previous step. Then add an additional 40µl of the DNase Reaction Buffer.

Mix DNase with buffer by gently inverting the tube a few times.

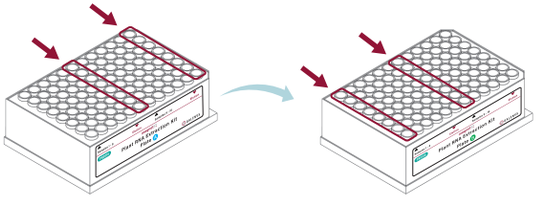


9 To the **RNA Extraction Kit Plate A** add 50µl of DNase Buffer to Elution Buffer (columns 6 & 12). Dispense directly into the elution buffer not on the walls. If there are any droplets of elution buffer left on the walls, slide them to the well bottom by gently shaking or tapping the plate on a solid surface.

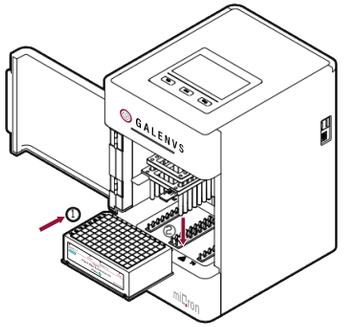


10 Gently shake **Plate A** for 10 seconds by hand and incubate at room temp for 20 mins.

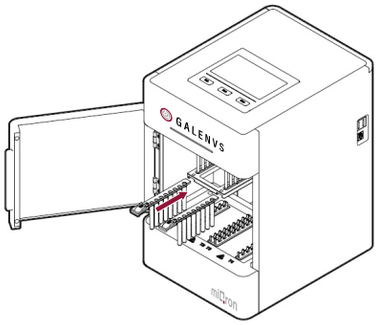
From the **RNA Extraction Kit Plate A** transfer 100uL of the Elution Buffer (columns 6 & 12) to each well (columns 1 & 7) of the **RNA Extraction Kit Plate B**



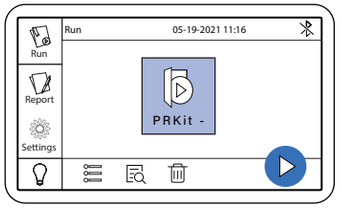
11 Place **Plate B** into the miQron, taking care that the label is facing outward.



12 Insert two combs.



13 Select **PRKit - Part B** and press ▶



When program is complete, remove plate from miQron and discard combs.

Columns 5 and 11 contain the purified RNA elution.