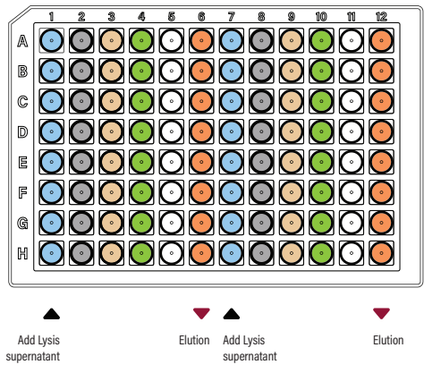
GALENVS

magnetiQ

Wastewater DNA/RNA Extraction Kit

Quick Start Guide



- Binding Buffer (600µl) — Columns 1 & 7
- Binding magnetic nanoparticles (30µl) — Columns 2 & 8
- Wash #1 Buffer (600µl) — Columns 3 & 9
- Wash #2 Buffer (600µl) — Columns 4 & 10
- Elution Buffer (100µl) — Columns 6 & 12

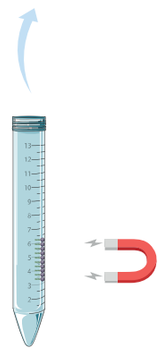
WWKit miQron protocol parameters

Step Name	Column	Volume (µl)	Time (sec)	Mixing Speed (1–10)	Dry Time (sec)	Magnet Capture Time (sec)	Temp
Bead Pickup	2 & 8	30	30	5	0	30	OFF
Binding	1 & 7	1000	300	5	0	150	OFF
Wash #1	3 & 9	600	60	6	0	60	OFF
Wash #2	4 & 10	600	60	6	60	60	OFF
Elution	6 & 12	100	300	7	0	270	65°C
Discard Comb	5 & 11	500		5			

1. Transfer 10ml of wastewater sample to 15ml conical tube. Add 100µl of Concentration Beads. Invert 5 times and incubate for 10 mins. Invert 3 more times at the 5 min mark.

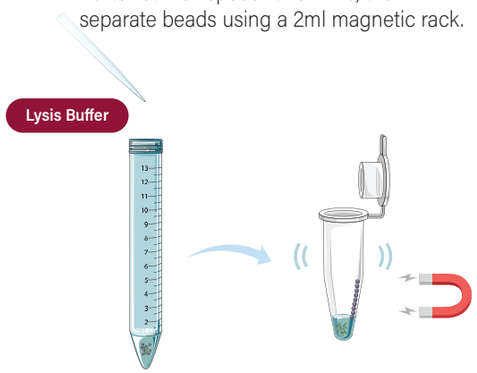
2. Place sample on 15ml magnetic rack to capture beads, then discard supernatant.

   Remove tube from magnetic rack.



3. Resuspend beads in 400µl of Lysis Buffer, then transfer to 2ml centrifuge tube.

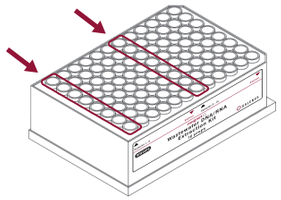
   Vortex at max speed for 5 mins, then separate beads using a 2ml magnetic rack.



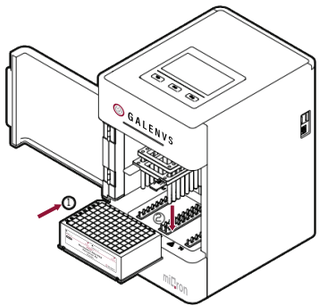
5. Remove the protective foil.

   Avoiding pellet, transfer up to 400µl of supernatant to the Binding Buffer wells (columns 1 & 7).

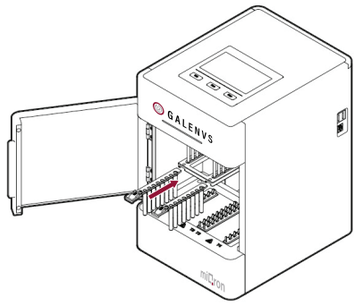
   You can add up to 16 samples.



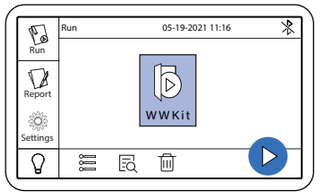
6. Place plate into the miQron, taking care that the label is facing outward.



7. Insert two combs.

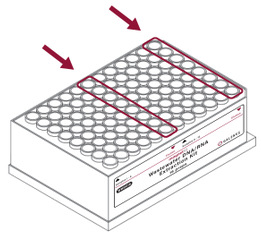


8. Select the WWKit protocol and press ▶



When program is complete, remove plate from miQron and discard combs.

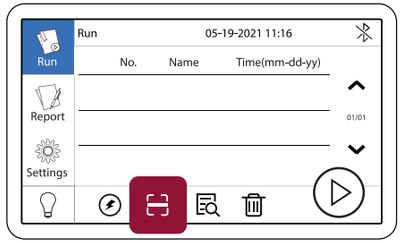
Columns 6 & 12 contain the purified DNA/RNA elution.



miQron

Protocol Update

To import the updated protocol into the miQron, press the **Scan Protocol** icon from the **Run** menu (protocol list view window). Use the scanner on the QR code below.



WWKIT PROTOCOL V1.1