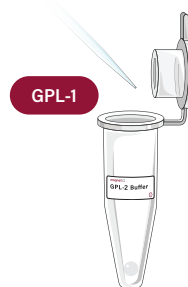


Quick Plasmid Miniprep Kit

Quick Start Guide

1 Add 4.4ml of GPL-1 solution to hydrate the GPL-2 tube containing lyophilized pellets. Ensure the prepared GPL-2 Buffer is mixed well by vortexing.

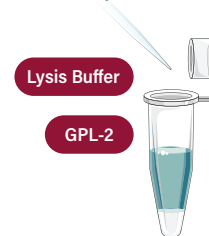
! Each GPL-2 tube provided contains 11 lyophilized pellets and is sufficient for 110 preps. The unused mixture should be kept at 4°C for storage and later use.



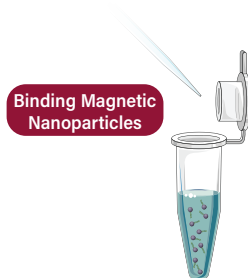
2 Pellet 1–10ml of bacterial cell sample. Resuspend bacterial pellet in 860µl of Lysis Buffer, then add 40µl of GPL-2 Buffer.

Mix gently by pipetting up-down until solution is homogenous. Incubate for 5 mins at room temperature.

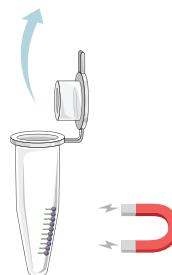
! For 10ml bacterial culture, add 80µl of GPL-2 Buffer for better lysis.



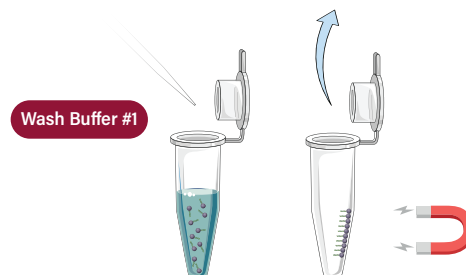
3 Add 40µl of Binding Magnetic Nanoparticles. Mix well by pipetting up-down (5–10x) until solution is homogenous. Incubate for 2 mins at room temperature.



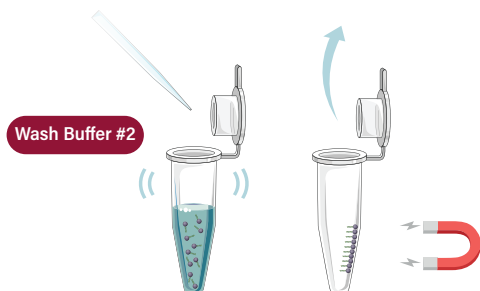
4 Place tube on magnetic rack to capture. Wait 1 min then discard supernatant.



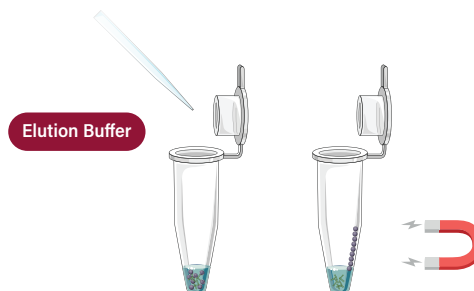
5 Remove tube from magnetic rack and resuspend in 600µl of Wash Buffer #1, and mix well by pipetting up-down (5–10x) until solution is homogenous. Return to magnetic rack for 1–2 mins, then discard supernatant.



6 Repeat wash with 600µl of Wash Buffer #2. Place tube on magnetic rack to capture. Wait 1 min then discard supernatant, and leave to dry for 2 mins.



7 Add 100µl of Elution Buffer to tube and mix well by pipetting up-down (10–15x) until solution is homogeneous. Incubate for 5 mins. Place tube on magnetic rack for 1 min.



8 Transfer pure plasmid DNA supernatant to a clean microfuge tube.

