

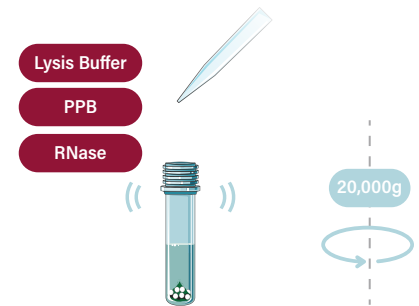
## Plant DNA Extraction Kit

## Quick Start Guide

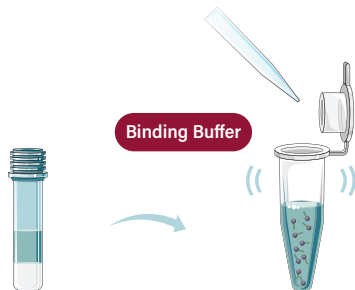
- 1 Add up to 50mg of ground fresh or dried plant leaves to the lysis bead tube provided.



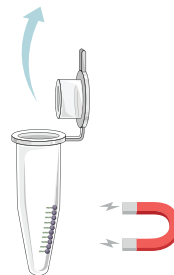
- 2 Add 600µl of Lysis Buffer, 60µl PPB and 5µl RNase.
- Mix for 3 mins using TissueLyser at max speed or vortex for 10 mins; then centrifuge at 20,000g for 2 mins.



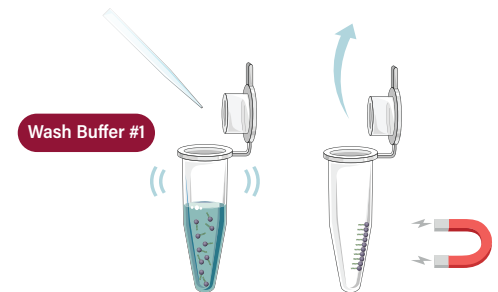
- 3 Avoiding pellet, transfer up to 300–400µl of supernatant to clean centrifuge tube.
- Add 1000µl of Binding Buffer, vortex for 10–20s, and wait 5 mins.



- 4 Place tube on magnetic rack to capture. Wait 1 min then discard supernatant.



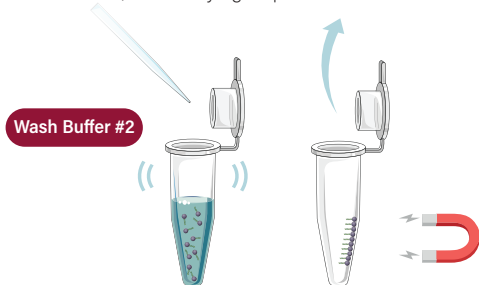
- 5 Add 600µl of Wash Buffer #1 to the tube and vortex for 10–20s.
- Wait 1 min then place tube on magnetic rack to capture. Wait 1 min then discard supernatant.



- 6 Add 600µl of Wash Buffer #2 to the tube and vortex for 10–20s. Place tube on magnetic rack to capture. Wait 1 min then discard supernatant.

Repeat step 6 twice.

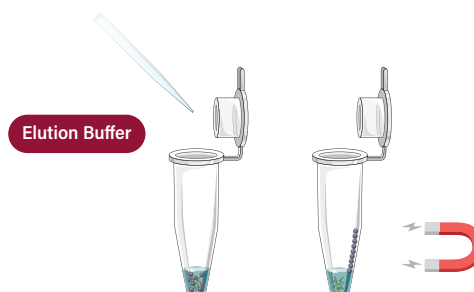
- ! Make sure to remove residual Wash #2 remaining in tube, or use drying step before elution.



- 7 Add 100µl of Elution Buffer to tube and mix briefly. Wait 1 min.

Place tube on magnetic rack to capture.

- ! For increased yield heat Elution Buffer at 60°C for 5 mins.



- 8 Wait 1 min then transfer supernatant to clean microfuge tube..

