GALENVS

magnetiC

Wastewater DNA/RNA Extraction Kit

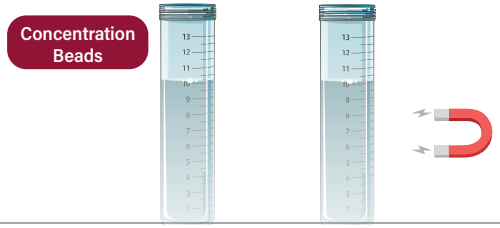
Quick Start Guide

1 Add 100µl of Concentration Beads (mix well before use) to 10ml wastewater sample, then invert 5 times to mix thoroughly.

Incubate for 10 mins. At the 5 min mark, invert 3 times to mix the beads.

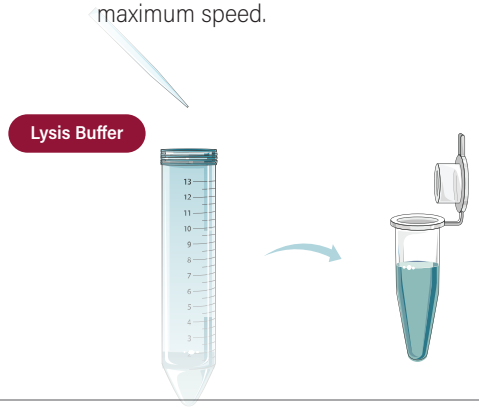
Place sample on 15ml magnetic rack to capture beads, then discard supernatant.

Note: Store the beads at 4°C.



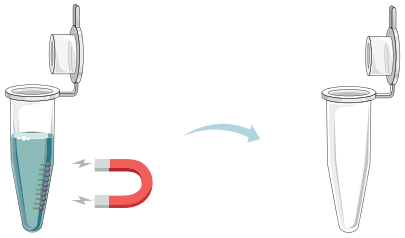
2 Add 400µl of Lysis Buffer. Resuspend the beads by pipetting up-down.

Transfer mixture to a clean 2ml centrifuge tube. Mix for 5 mins using vortexer at maximum speed.



3 Use a 2ml magnetic rack to capture the beads.

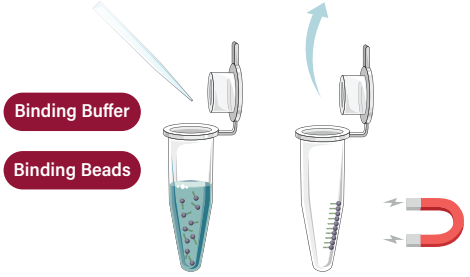
Avoiding the pellet, transfer up to 400µl of the supernatant into a clean 2ml microcentrifuge tube.



4 Add 600µl of Binding Buffer.

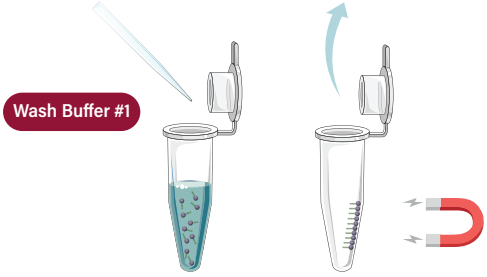
Add 30µl of Binding Beads. Vortex for 10-20s, and incubate for 5 mins.

Place tube on magnetic rack to capture, wait 1 min then discard supernatant.



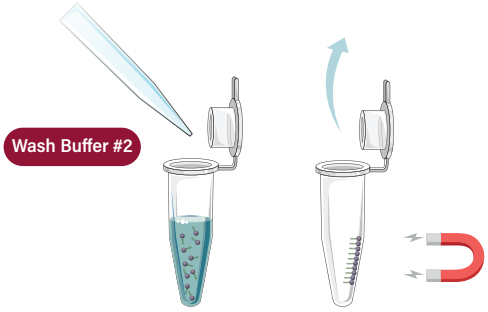
5 Add 600µl of Wash Buffer #1 to the tube and vortex for 10–20s.

Wait 1 min then place the tube on the magnetic rack to capture. Wait 1 min then discard supernatant.



6 Add 600µl of Wash Buffer #2 to the tube and vortex for 10–20s.

Place the tube on a magnetic rack to capture. Wait 1 min then discard supernatant.



7 Add 100µl of Elution buffer to tube and mix briefly.

Incubate at 65°C for 10 mins.

Place the tube on a magnetic rack to capture.

8 Wait 1 min then transfer supernatant to clean 2ml microcentrifuge tube.

Note: For increased yield perform elution twice with 50µl of buffer.