GALENVS

magnetiQ

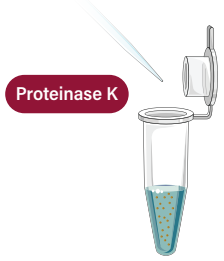
Blood & Cell DNA Extraction Kit

Quick Start Guide

1 **In 1.5ml microfuge tube, add 20µl of Proteinase K.** Then add 100µl of whole blood[1] sample or 100µl of cell suspension[2] in 1x PBS.

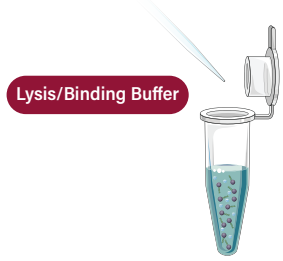
[1] *For 200µl of whole blood, use 600µl of Lysis/Binding Buffer in step 2*

[2] *Up to 1x10⁶ cells in 100µl*

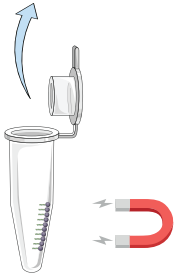


2 **Add 400µl of Lysis/Binding Buffer** and mix well by pipetting up-down 20x.

Incubate 5 mins at room temp (20-25°C) to allow for lysis and DNA binding.

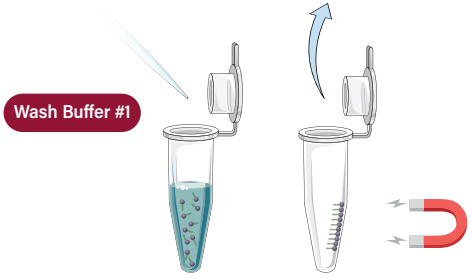


3 **Place tube on magnetic rack for 5 mins** to capture DNA-bead complex, then discard supernatant.

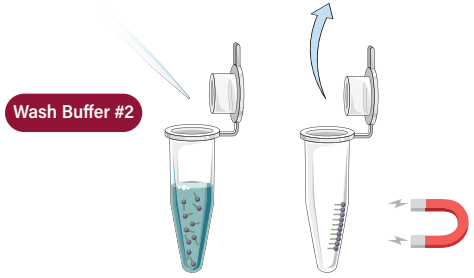


4 Remove tube from magnetic rack and **resuspend DNA–bead complex in 600µl of Wash Buffer #1**. Mix well by pipetting up-down 10–15x.

Return to magnetic rack for 5 mins, then discard supernatant.



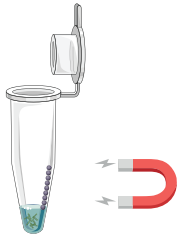
5 **Repeat wash with 600µl of Wash Buffer #2**, return to magnetic rack for 1–2 mins, then discard supernatant and leave to dry for 1 min.



6 Remove tube from magnetic rack and **resuspend DNA–bead complex in 50–100µl of Elution Buffer*.** Mix well by pipetting up-down 15–20x to elute DNA from beads and let stand for 1–2 mins.

* *Recommended elution is 50–100µl for blood sample and 100µl per 1x10⁶ cell suspension sample*

7 **Place tube on magnetic rack to capture beads (~1–2 mins).**



8 Transfer the eluted DNA solution (supernatant) to a clean microfuge tube.